Evaluation of the Interrater Reliability of Sonographic Measurements of Muscle Thickness of 38 Piriformis Muscles in 19 Patients with Piriformis Syndrome

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Background:
The piriformis muscle is a flat superficial muscle of the deep gluteal muscles that externally rotates the hip. Ultrasound is widely used to identify the piriformis muscle, especially for guidance of the needle during injections; however, its diagnostic use has recently gained popularity. The operator-dependent nature of ultrasound requires demonstration of reliability between operators. This study aimed to evaluate interrater reliability of sonographic measurements of muscle thickness of 38 piriformis muscles in 19 patients with piriformis syndrome.

Material/Methods:
An ultrasound transducer was placed transversely on the sacral spinous process and moved caudo-laterally until the piriformis muscle was visualized under the gluteus maximus while patients were lying in prone position. The thickness of piriformis muscle was measured with a 2 to 5-MHz broadband curvilinear transducer in 3 regions (thickest regions of muscle over the ilium, near the greater trochanter, and near the sacrum). The interrater reliability of measurements of 2 examiners who were blinded to each other's measurements was assessed by intraclass correlation coefficient.

Results:
In total, 114 samples from 38 piriformis muscles of 19 patients with a diagnosis of piriformis syndrome were evaluated by 2 raters in this study. The median (interquartile range) patient age was 41 (15) years. Intraclass correlation coefficient value for overall thickness measurements of piriformis muscle was 0.836. Intraclass correlation coefficient values for 3 different regions were over the ilium, near the greater trochanter, and near the sacrum were 0.777, 0.883, and 0.811, respectively.

Conclusions:
Ultrasound measurement of piriformis muscle thickness has good interrater reliability.

Keywords: Piriformis Muscle Syndrome • Reproducibility of Results • Diagnostic Imaging • Ultrasonography

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Introduction

The piriformis muscle is a flat, pear-shaped external rotator of the hip, located deep in the gluteal region. The sciatic nerve exits the pelvis distal to this muscle except for some anatomical variations where the entire sciatic nerve or a division of it pass through or is proximal to the piriformis muscle [1]. The piriformis muscle has gained clinical significance in deep gluteal or extra-spinal sciatica pain syndrome owing to its close proximity to the sciatic nerve [2,3].

The piriformis muscle is not the sole muscle playing a role in deep gluteal pain [4], and “deep gluteal syndrome” is a more appropriate term to explain this clinical entity [5]; however, the term “piriformis syndrome” is still widely used today. This clinical condition is a constellation of symptoms, including buttock pain that can radiate down to the leg, and can be accompanied by sensory symptoms in the leg [3,6].

An irritated or inflamed piriformis muscle leading to the irritation of the sciatica nerve anterior to the piriformis muscle is thought to cause buttock and radiating leg pain in piriformis syndrome [6]. On the other hand, a hypertrophic, tight, and tender piriformis muscle might be responsible for the myofascial pain radiating down the leg without sciatic nerve compression [7]. On physical examination, palpation over the piriformis muscle and several specific maneuvers, such as flexion, abduction, internal rotation (FAIR), and pace (resisted abduction and external rotation of the thigh), provoke pain [6]. Diagnosis depends mainly on symptoms and clinical findings. Studies regarding radiological data showing changes in the piriformis muscle are evolving [7-9].

A hypertrophic piriformis muscle can be observed on magnetic resonance imaging (MRI) [3,8]. MRI and/or computerized tomography (CT) imaging exhibited pathologies related to the piriformis muscle in 64% of 116 patients with clinically suspected piriformis syndrome, and piriformis muscle enlargement was the most common imaging finding [9]. The authors suggest that cross sectional imaging of the piriformis muscle can alter treatment planning. Their study population mainly consisted of patients with trauma, infarction, infection, tumor, or other space-occupying lesions, which may explain the high rate of abnormal imaging findings.

Ultrasound is widely used in the identification of the piriformis muscle, especially for guidance of the needle during injections [10], and the diagnostic role of ultrasound has recently gained popularity in the research field [11-16]. On ultrasound, thickness of the piriformis muscle was measured as greater in patients with piriformis syndrome, by 0.4 cm and 0.5 cm, than the asymptomatic side of the same patient and healthy people, respectively [12,14]. Its low cost, easy accessibility, and absence of ionizing radiation enhances the value of ultrasound. However, its highly operator-dependent nature requires demonstration of reliability between different operators [10]. Present studies have not investigated reliability. Therefore, in this study, we aimed to evaluate the interrater reliability of sonographic measurements of piriformis muscle thickness in patients with piriformis syndrome.

Material and Methods

The study was approved by the local ethics committee (2018.053.IRB1.009). All participants provided consent prior to the study, which was conducted in the Physical Medicine and Rehabilitation Unit, Bakirkoy Dr Sadi Konuk Training and Research Hospital, Bakirkoy, Turkey, between January 2020 and March 2022.

Participants

Nineteen patients (15 women, 4 men) diagnosed with piriformis syndrome were included in the study. The inclusion criteria were patients aged >18 years with buttock pain and positive physical examination findings of piriformis syndrome, including tenderness upon palpation over the piriformis muscle, a positive FAIR test, and a positive pace maneuver [3,6,7]. Exclusion criteria were radiculopathy, low back pain, history of lower extremity or pelvic surgery, inability to walk, malignancy, and rheumatologic or neurologic disorders. Age, sex, height, and weight of all patients were recorded, and body mass index was calculated.

Data Collection

A Hitachi Noblus (Hitachi Aloka Medical America, Inc) ultrasonic imaging system was used, with a 2 to 5-MHz broadband curvilinear (60 mm radiant) transducer. Device settings regarding image depth and gain were set according to the thickness of the overlying tissues of each patient to accurately visualize the deeply located piriformis muscle.

Participants were asked to lie in the prone position on the examination table with the gluteal regions undressed. Lower extremities were in the neutral position. The curvilinear transducer was placed directly onto the skin using sufficient transmission gel. The scanning protocol was started with placement of the ultrasound transducer transversely over the midline on the sacral spinous process, which was moved laterally to the posterior superior iliac spine and sacroiliac joint of the imaged side. Then, the transducer was moved caudally toward the greater sciatic foramen [17]. After visualizing the piriformis muscle under the gluteus maximus, the piriformis muscle was confirmed by demonstrating it got stretched with
passive internal rotation of the hip with the knee flexed in 90 degrees [16]. Then, the lateral end of the transducer was slightly rotated along the longitudinal axis of the piriformis muscle (Figure 1). Three measurements of piriformis muscle thickness were performed: thickest parts of piriformis muscle over the ilium, on the lateral side of the ilium near the greater trochanter, and on the medial side of the ilium, near the sacrum (Figure 2). Muscle thicknesses on longitudinal images were measured via the “distance” measurement function of the device. Measurements were performed in both extremities while the hip joint was in a neutral position.

Interrater Reliability

Two physiatrists, with intermediate to advanced musculoskeletal ultrasound skills, performed the ultrasound measurements. Prior to the study, the examiners conducted a pilot ultrasound study on 4 volunteers to standardize the scanning planes and measurement sites.

For interrater reliability, one of the examiners performed the ultrasound evaluation after the other on the same day. The examiners were blinded to each other’s examinations and measurements.

Statistical Analysis

The power analysis was calculated using the online sample size tool [18] at the 95% power level and 5% statistical significance level. Under the assumptions of minimum acceptable reliability ($p_r$) of 0.70 and expected reliability ($p_e$) of 0.84 with 2 raters (k), it was calculated that 105 samples would be sufficient. In our study, each rater measured a total of 114 samples (3 measurements for each muscle, summing 3×38 measurements), reaching this sample size.

SPSS version 22.0 for Windows (IBM Corp, Armonk, NY, USA) was used for statistical analysis. The normality of the data distribution was analyzed using the Shapiro-Wilk test. Continuous variables with a normal distribution are presented as mean±standard deviation and continuous variables that were not normally distributed are presented as median (interquartile range). Categorical variables are presented as numbers. To compare muscle thickness measurements between the symptomatic and normal sides, the Wilcoxon signed-rank test was used. The interrater reliability of ultrasound measurements was assessed by the intraclass correlation coefficient analysis with the absolute agreement definition and 2-way mixed-effects model. Intraclass correlation coefficient values below 0.5, from 0.5 to 0.74, 0.75 to 0.89, and greater than 0.90 were defined as poor, moderate, good, and excellent reliability, respectively [19].

Results

Demographic Characteristics

In total, 114 samples from 38 piriformis muscles of 19 patients were evaluated by 2 raters in this study. The median (interquartile range) age was 41 (15) years (min-max, 29-79). Mean height, weight, and body mass index values were 1.65±0.06 m (min-max, 1.50-1.80), 66.9±9.5 kg (min-max, 51-92) and 24.68±3.17 kg/m$^2$ (min-max, 18.73-32.21), respectively. Two patients were left-handed.
The intraclass correlation coefficient value for overall thickness measurements of piriformis muscle was 0.836. The intraclass correlation coefficient values in 3 regions of piriformis muscle over the ilium, near the greater trochanter, and near the sacrum were 0.777, 0.883, and 0.811, respectively (Table 2).

When the data were separated as symptomatic and asymptomatic side, there were no differences in the mean values of muscle thickness between the symptomatic and normal sides at 3 measurement sites (Table 3). The intraclass correlation coefficient value for overall thickness measurements of piriformis muscle was 0.877 (95% CI: 0.706-0.949, P<0.001) and 0.896 (95% CI: 0.616-0.968, P<0.001), for symptomatic and asymptomatic sides, respectively. Individual values for muscle thickness measurements of each examiner for dominant and non-dominant sides are given in Table 1.

Table 1. Individual measurement values of piriformis muscle thickness measurements from different measurement sites in dominant and non-dominant sides of patients with piriformis syndrome by each examiner.

<table>
<thead>
<tr>
<th>Measurement site</th>
<th>Mean thickness of piriformis muscle (mm)</th>
<th>Examiner 1</th>
<th>Examiner 2</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>mean±SD (min-max)</td>
<td>Dominant</td>
<td>Nondominant</td>
</tr>
<tr>
<td>Near greater trochanter</td>
<td>11.0±2.7 (6.0-17.9)</td>
<td>10.9±3.0 (7.5-19.1)</td>
<td>10.4±2.8 (6.4-16.7)</td>
</tr>
<tr>
<td>Over ilium</td>
<td>11.8±2.4 (7.8-16.6)</td>
<td>11.8±2.4 (8.5-16.9)</td>
<td>12.2±2.4 (8.5-16.6)</td>
</tr>
<tr>
<td>Near sacrum</td>
<td>12.3±2.9 (8.1-19.3)</td>
<td>12.4±3.6 (8.0-22.4)</td>
<td>12.8±3.1 (8.4-21.8)</td>
</tr>
</tbody>
</table>

**Table 2.** Interrater reliability of muscle thickness measurements between 2 examiners.

<table>
<thead>
<tr>
<th>Measurement site</th>
<th>Intraclass correlation</th>
<th>95% Confidence interval</th>
<th>F test with True 0</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Lower bound</td>
<td>Upper bound</td>
</tr>
<tr>
<td>Near greater trochanter</td>
<td>0.883</td>
<td>0.769</td>
<td>0.940</td>
</tr>
<tr>
<td>Over ilium</td>
<td>0.777</td>
<td>0.570</td>
<td>0.885</td>
</tr>
<tr>
<td>Near sacrum</td>
<td>0.811</td>
<td>0.632</td>
<td>0.903</td>
</tr>
<tr>
<td>Overall</td>
<td>0.836</td>
<td>0.761</td>
<td>0.888</td>
</tr>
</tbody>
</table>

Table 3. Individual measurement values of piriformis muscle thickness measurements from different measurement sites in symptomatic and normal sides of patients with piriformis syndrome by each examiner.

<table>
<thead>
<tr>
<th>Measurement site</th>
<th>Mean thickness of piriformis muscle (mm)</th>
<th>Examiner 1</th>
<th>Examiner 2</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>mean±SD (min-max)</td>
<td>Symptomatic</td>
<td>Normal</td>
</tr>
<tr>
<td>Near greater trochanter</td>
<td>10.9±2.6 (7.5-17.9)</td>
<td>10.9±3.2 (6.0-19.1)</td>
<td>0.414</td>
</tr>
<tr>
<td>Over ilium</td>
<td>12.0±2.4 (8.5-16.6)</td>
<td>11.5±2.3 (7.8-16.9)</td>
<td>0.510</td>
</tr>
<tr>
<td>Near sacrum</td>
<td>12.6±3.7 (8.0-22.4)</td>
<td>12.0±2.4 (9.2-16.6)</td>
<td>0.701</td>
</tr>
</tbody>
</table>

mm – millimeter; min – minimum; max – maximum; SD – standard deviation.

Piriformis muscle thickness measurements for each examiner for dominant and non-dominant sides are given in Table 1. The intraclass correlation coefficient value for overall thickness measurements of piriformis muscle was 0.836. The intraclass correlation coefficient values in 3 regions of piriformis muscle over the ilium, near the greater trochanter, and near the sacrum were 0.777, 0.883, and 0.811, respectively (Table 2).
thickness at 3 different points of piriformis in the symptomatic and normal side measured by examiner 1 are presented in Figures 3 and 4, respectively.

**Discussion**

This study demonstrated that ultrasound measurement of piriformis muscle thickness had good reliability when 2 different experienced examiners performed the measurements. The interrater reliability is clinically important during the diagnosis and follow-up of patients in both daily practice and research studies. Recent studies suggested morphological changes such as increased thickness [10-15], enlarged cross sectional area [11], change in echo-intensity [11,12] and elastography strain ratio [15,16] of the piriformis muscle or increased diameter of the sciatic nerve [12] in the symptomatic side of patients with piriformis syndrome. However, none of the studies investigated interrater reliability.

Piriformis muscle thickness measured by ultrasound is suggested to be able to discriminate between the symptomatic and asymptomatic sides in piriformis syndrome, with different cut-off values in different studies, such as 18.15 mm (sensitivity, 72%; specificity, 74%) [12], 17.85 mm (sensitivity, 60%; specificity, 95%) [16], and 9.95 mm (sensitivity, 95%; specificity, 88%) [13]. Two studies suggested similar values nearly twice that of the other study, leading to questions about the use of an absolute value for a cut-off point.

Measurement sites and positions differ among these studies. Siahaan et al measured thickness from the medial part of the tip of ilium parallel to the longitudinal plane at the sciatic notch while the leg of the patient was in 45-degree abduction [13]. This region is somewhat similar to over ilium measurement in our study; however, the position of the patient’s leg was neutral during our measurement. Other studies perform the measurement from the thickest portion of the muscle [12,14]. This corresponds most probably to the near sacrum site in our study. However, as shown in Figures 3 and 4, the thickest site of the piriformis might be any of the 3 measurement sites. This finding might be explained by the various shapes of the piriformis muscle. Since we demonstrated that overall measurement sites are reliable, any site can be used to measure thickness. However, to compare different measurements, it is essential to note the location of measurement in the records.

A scanning protocol for the piriformis muscle dynamically evaluating and confirming its visualization in healthy participants was defined by Battaglia et al, and was similar to our scanning protocol [17]. Passive internal and external rotation of the hip during scanning helps to distinguish the borders of the deeply located external rotators from the surrounding muscles. In that study, 2 ultrasonographers rated the diagnostic value of dynamic ultrasound in the pathologies of piriformis muscle as “likely”, but they neither performed a reliability study nor measured the thickness of the muscle. Broadhurst et al also investigated the morphology of the piriformis muscle by
ultrasound, but only qualitatively [20]. Among a total of 27 patients with buttock pain, they found enlarged muscle in 15 patients, scarred and fibrotic muscle in 3 patients, and normal morphology in 9 patients. However, they did not clearly explain their scanning technique and did not make any quantitative measurement.

MRI was used to measure the cross-sectional area of the piriformis muscle by Grimald et al in advanced degenerative hip disease. They reported similar muscle volumes in the degenerated leg by 14% and high interrater reliability (intraclass correlation coefficient: 0.985) [21]. In low back pain with leg pain, piriformis muscle atrophy was detected on the symptomatic side on MRI [22]. Muscle volume measurements were performed by 3-dimensional manual segmentation of MR images and they reported high inter-rater reliability. This technique is highly costly and time-consuming. Eastlack et al [23] studied the intra- and interrater reliability of piriformis muscle assessment in axial T1-weighted images in 134 patients with pelvic dysfunction and back or buttock pain. They found that intra- and interrater reliabilities were moderate to good (kappa values were 0.39-0.60 and 0.74-0.82 for inter- and intraobserver, respectively). They categorized piriformis muscle asymmetry qualitatively. Approximately one-third of the patients had piriformis muscle asymmetry, with hypertrophy more common than atrophy.

Russell et al [24] measured the thickness of the piriformis muscle in T1-weighted MRI in 100 patients, 77% of whom had back or buttock pain. They measured from only a single site (approximately 1-2 cm inferior to the sacroiliac joint), and the mean of their measurements was 1.9 cm, ranging from 0.8 to 3.2 cm. They measured the depth of the piriformis muscle from its posterior to anterior border in axial images. However, in the present ultrasonographic study, we measured the thickness of the piriformis muscle from its superior to inferior border. Our measurements could be compared with the values measured in coronal MR images.

In a MR neurography imaging study, Filler et al measured the piriformis muscle preoperatively in nearly 50 patients with piriformis syndrome, who had good to excellent outcomes after piriformis surgery. They observed ipsilateral piriformis muscle hypertrophy in 38.5% and atrophy in 15% in preoperative MR images [25]. Muscle asymmetry alone had lower specificity (66%) and sensitivity (46%) values in identifying piriformis syndrome than did including the asymmetry observed in hyperintensity of the sciatic nerve (specificity and sensitivity, 93% and 64%, respectively).

Haladaj et al evaluated anthropometric features of the piriformis muscle and sciatic nerve in 30 limbs of cadavers and reported the thickness of piriformis muscle as 31.6 mm in cadavers with normal muscle morphology [26]. They did not specify the location where they measured the thickness of piriformis muscle.

Park et al measured piriformis muscle thickness at 3 sites at the midsciatric notch level with CT scanning in 60 patients with no evidence of piriformis syndrome [27]. This site corresponds to the proximal part of the muscle that we measured near the sacrum in our study; however, they measured the depth of the piriformis muscle, similar to the MRI study by Russell et al.

Measurements by MRI or CT imaging mainly involve the depth size of the piriformis muscle in a single region. Thickness can be measured from coronal images. Ionizing radiation, low accessibility, and high economic cost are the disadvantages for these imaging studies. Since MRI is not an examiner-dependent technique, with excellent soft tissue resolution, it might serve as a comparison in validity studies of ultrasound. However, none of the recent studies mentioned above compared the ultrasound findings with that of MRI. One study measured the thickness and cross-sectional area of piriformis muscle by both ultrasound and MRI; however, ultrasonographic measurement results were not validated with MRI measurements [10].

One of the limitations of the present study is that intrarater reliability was not performed. This was not included in our study design not to invite the participants to the hospital in another day. Another limitation is that we did not perform a sample size analysis for comparing muscle thickness measurements between symptomatic and normal sides; therefore, our sample size is small for interpreting our findings in relation to the relevant literature.

The strength of this study is that this is the first interrater reliability study for measuring piriformis muscle thickness. The piriformis muscle was measured from 3 different sites, different from other studies.

Conclusions

Ultrasound measurement of piriformis muscle thickness was shown to have a good interrater reliability between 2 physiatrists who are experienced in musculoskeletal ultrasound.

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References:


